Hypoxia Signaling to Genes

Significance in Alzheimer's Disease

Nicolas G. Bazan,¹ Ricardo Palacios-Pelaez,² and Walter J. Lukiw,*,²

¹Neuroscience Center and Department of Ophthalmology, Louisiana State University Health Sciences Center, New Orleans, Louisiana 70112-2272 USA; ²Diater Laboratorios, San Martin de la Vega-Madrid, Spain

Abstract

Aberrations in neural signaling, converging to and diverging from oxidative metabolism and blood supply, contribute to the initiation and maintenance of inflammatory responses, neuronal degeneration, and age-related cognitive decline in Alzheimer's disease (AD). Hypoxia/ischemia triggers phospholipase A2, leading to the accumulation of free arachidonic and docosahexaenoic acids (AA, DHA), as well as that of lysophospholipids. Some of these bioactive lipid messengers in turn give rise to several downstream lipid messengers, such as platelet-activating factor (PAF) and ecosanoids (prostaglandins and leukotrienes). Eicosanoid synthesis is highly regulated in hypoxia and in reperfusion subsequent to ischemia. As one of the consequences, mitochondrial function is disrupted and reactive oxygen species (ROS) both contribute to the expansion of cellular inflammatory responses and reduce the expression of genes required to maintain synaptic structure and function. On the other hand, pro-inflammatory genes are up-regulated. One of these, the inducible cyclooxygenase-2 (COX-2), along with oxygen-starved mitochondria, comprise the major sources of ROS in the brain during hypoxia, ischemia, and reperfusion. One outcome is a sustained metabolic stress that drives progressive dysfunction, apoptosis and/or necrosis, and brain cell death. How hypoxia modulates oxygen-sensitive gene expression is not well understood. Pro-inflammatory gene families that contribute to neurodegeneration are transiently activated in part by the heterodimeric oxygen-sensitive DNA-binding proteins nuclear factor for kappa B (NF-κB) and hypoxia-inducible factor-alpha (HIF-1α). Here the authors summarize current studies supporting the hypothesis that synaptically-derived lipid messengers play significant roles in ischemic stroke and that hypoxia is an important contributor to the onset and progression of AD neurodegeneration.

Index Entries: Alzheimer's disease; beta-amyloid; brain transcription; cyclooxygenase-2 (COX-2); HIF-1α; hypoxia; inflammatory signaling; ischemia; gene expression; presenilin-1 (PS1); presenilin-2 (PS2); reactive oxygen species (ROS).

^{*}Author to whom all correspondence and reprint requests should be addressed. E-mail: wlukiw@lsuhsc.edu

Introduction

Neurons are highly vulnerable to impairments in oxygen homeostasis because of their singular dependence on oxygen. The human brain averages just 2% of body mass, yet utilizes 15% of cardiac output and 20% of respiratory oxygen uptake. The brain undergoes damage and neural dysfunction after only a few minutes of oxygen deprivation; if oxygen deprivation lasts longer than 5 min, apoptotic and/or necrotic cell death occurs. In neural cells in primary culture and in the hippocampus in in vivo models, both cyclooxygenase-2 (COX-2) and presenilin-1 (PS1) are induced after only 5 min of hypoxia/ischemia (1–8). Overexpression of these pro-inflammatory genes appears to redirect cellular fate toward intracellular signaling dysfunction and apoptotic cell-death (9–21). For example, hypoxia/ischemia enhances COX-2 expression in neurons of the cerebral cortex and hippocampus, and as a result, prostaglandins (PG) and reactive oxygen species (ROS), principally the peroxides and hydroxides O_2 -, OH-, and H_2O_2 , are generated. PG overproduction and reactive oxygen species promote transient and concentration-dependent neuronal signaling dysfunction. After brief periods of hypoxia, re-establishment of normoxia is accompanied by a surge of ROS and inflammatory gene induction (8,10,15,18–22). enhanced physiologic signaling initiates and amplifies brain-specific inflammatory responses to hypoxia, leading to the robust synthesis of cytokines, PGs, eicosanoids, related lipid mediators, and ROS (18,19,23-25). Cell cultures and transgenic models suggest interactive relationships between hypoxia, microglial activation, neuro-inflammation, reduced neuronal function, and apoptosis and/or necrosis (19–21). Indeed, hypoxic injury triggers COX-2-mediated arachidonic acid (AA) metabolite synthesis, which in turn contributes to altered metabolism and AD neuropathology (8,10,15,26,27).

Epidemiology of Alzheimer's Disease

AD is a complex, progressive degeneration of the neurons of the neocortex, characterized

by memory loss and deterioration of higher cognitive function. Aging is the most significant risk factor for AD; however, the exact sequence of events that initiates this disorder remains unclear. Both AD prevalence and incidence double every 5 yr after the age of 65 yr. AD currently affects about 8% of the US population aged 65 yr and over and increased cognitive deficits strongly correlate with AD mortality. Demography indicates that the rapidly aging US population will soon incur massive increases in the number of AD cases that will overwhelm our healthcare system and exceed our ability to adequately care for people afflicted with this devastating disorder of the mind (12–22,26,28–30).

Besides aging, an intricate maze of factors contributes to the development and progression of AD. Epidemiologic studies indicate that mutations in gene-coding regions, susceptibility polymorphisms in gene-control regions, traumatic head injury, environmental factors involving zinc and/or aluminum, or other environmental neurotoxins, cerebrovascular disease, and ischemia and/or hypoxia, are factors for AD development (30–37). Classically, the density of neurofibrillary tangles (NFT) and peptides derived from beta-amyloid precursor protein (βAPP) have been correlated with AD progression. However, recent evidence suggests that neuronal atrophy and disintegration of synaptic contacts are more relevant pathologic markers (9,38,39). Molecular genetic evidence indicates considerable heterogeneity in AD. For example, while a small number of familial AD cases exhibit linkage to mutations in the βAPP, PS1, and PS2 genes, which encode integral membrane proteins, or the presence of the apolipoprotein E-epsilon 4 (ApoE4) allele, which encodes a blood-borne lipoprotein carrier, the vast majority of AD cases are of sporadic or unknown origin (26,29,37,40-43). Moreover, while specific βAPP, PS1, PS2, and ApoE genotypes are associated with different relative risks and age-of-onset distributions for AD etiopathology, genetic variation also influences important aspects of clinical AD phenotype, such as severity, symptomatology, and disease duration (24–26,40–43).

Pathophysiology of Alzheimer's Disease: Amyloid and Reactive Oxygen Species

AD pathophysiology involves proliferation of astrocytes, gliosis and microglial activation, alterations in neuronal shape, cytoskeletal abnormalities, synaptic and neurotransmitter deficits, transcriptional, translational, and protein-processing defects, and progressive atrophy and loss of large neurons. AD was originally characterized by the presence of agyrophilic brain lesions that include abnormally hyper-phosphorylated forms of the microtubule-associated protein tau, which forms the core of intraneuronal NFT, and also by aggregates of insoluble extracellular beta-amyloid (A β) protein, derived from tandem β - and γ secretase cleavage of βAPP. Interestingly, autosomal dominant mutations in familial AD in the βAPP gene-coding region associated with the β - and γ -secretase cleavage sites lead to increases in Aβ peptide generation, and in particular that of the highly toxic A β 42 variant. Heterogeneous γ-secretase cleavage sites produce an array of ragged AB fragments, from 39–43 amino acid residues in length; Aβ peptides and especially the A β 42 variant appear to be specifically detrimental to neural function. Firstly, Aβ peptides directly induce ROS generation, thereby activating various kinases (such as p38, PKC, ERK2, Src, RTK) via the MAPkinase and related pathways, leading to aberrant intra-cytoplasmic phosphorylation and clumping and/or crosslinking of neural intermediate filaments (39–41,44). Secondly, ROS potently induce tau glycation and promote lipid peroxidation, leading to disruption in neural membrane physiology (15,24,27,37,43). In addition to deregulating intracellular protein kinase signaling, both NFT and Aβ aggregates contribute to disruption and extracellular disorganization of synaptic circuitry in the association hippocampus, neocortex, and basal forebrain cholinergic system, leading to impairments in inter-neural signaling and cognitive

dysfunction (35–38). Moreover, disturbances in βAPP gene expression may have consequences in the expression of other AD modifier genes, such as PS1 and PS2 (41–43). For example, animals doubly transgenic for mutant βAPP and PS1 show altered βAPP processing and accelerated AB formation, indicating contributions to AD pathology from at least two chromosomally unlinked genes. However, deficiency of other features of AD in doubly transgenic animals, such as absence of NFT and neuronal loss, suggests that additional factors are involved in modifying AD genotype (18,24,45). Widespread dysregulation in gene transcription supports the idea of alterations in entire families of brain-specific genes in both familial and sporadic AD. Deficits in the expression of neurotrophic factors, transcription factors (TFs), antioxidant enzymes, and cytoskeletal and synaptic components are all implicated in AD (9,38,39,46-48), as are the regulatory mechanisms that control transcription from these genes (8,48–51). Broad-spectrum DNA array analyses further corroborate that selective deregulation of brain RNA-message output is a consistent feature of AD, and that while the majority of gene transcripts are down-regulated, certain RNA messages encoding stress and inflammatory mediators exhibit marked overexpression (8,52–55). In addition, epidemiologic evidence indicates that antiinflammatory compounds, such as glucocorticoids and non-steroidal anti-inflammatory drugs (NSAIDs) ameliorate, at least partially, AD progression (55–58). COX-2, the key proinflammatory gene encoding an inducible oxidoreductase enriched in brain neurons, is consistently up-regulated both at the RNAmessage and protein level in the AD-affected neocortex and hippocampus, again implicating the activation of ROS-mediated signaling and inflammatory pathways in AD etiopathology (1,2,8,55,58-60).

Ischemia and Hypoxia as Risk Factors

Cerebral ischemia (restriction in blood supply) and hypoxia (decreased oxygen availabil-

ity) are potent inducers of Alzheimer-type change (28,43,61). Substantial evidence indicates that increased ROS are associated with aging and age-related degenerative disorders not only in AD, but also in Parkinson's disease, atherosclerosis, arthritis, and stroke (62). Besides selective neocortical neuron-death and progressive Aβ-deposition, AD neuropathology is further characterized by microglialmediated cytokine signaling. Interestingly, after 24 h of exposure, Aβ can elicit a greater reduction in neuroblastoma cell-viability and a greater increase in ROS than can glutamate (44). Indeed, hippocampal neurons appear to be exceptionally sensitive to gene-expression changes triggered by cytokines such as IL-1\beta, toxic peptides such as Aβ42, and hypoxia, and there appear to be cumulative and synergistic effects of these factors during aging (8,55). Postmortem examination of asphyxiated infants who have died from hypoxia has shown that the hippocampus was the brain locus where neuronal degenerative changes were most clearly observed, and these changes included astrogliosis and the appearance of stellate glia and AD-type astrocytic changes (4,7,61–63). Under nonlethal clinical conditions, early oxygen deprivation during human brain development may therefore predispose neural tissues to various subsequent events in endothelial cells and neurons during the perinatal period, with progressive metabolic dysfunction as the brain ages (8,13–15,20,63,64). Ischemic episodes later in life, such as repeated transient ischemic events, neurotrauma, or stroke, may also predispose the brain to ADdevelopment, and clinical data support this concept. The authors have tested these ideas in experimental models and portions of these results have been published (8,15,53).

Hypoxia and Nuclear Factor for Kappa B

Originally described as a novel *trans*acting nuclear protein that binds to the promoter of the immunoglobulin kappa light chain gene in human B cells (65–70), the significance of NF-κB has expanded to a more general role as an early

and rapid-response, ROS-sensitive regulator of genes involved in apoptosis, inflammation and the immune response, signal transduction, synaptic transmission, and neuronal plasticity (65-75). In the resting cytoplasm, NF- κ B exists predominantly as a p50-p65 heterotypic dimer electrostatically complexed with $I\kappa$ -B inhibitory protein; however, upon physiologic or pharmacologic induction, Ik-B is serine-phosphorylated, ubiquitinated, and degraded by the proteasome complex (72,73). This enables NF-κB to undergo translocation to the nucleus, where it binds target-DNA-consensus sequences homologous to 5'-GGGGACTTTCCC-3' in NF-κB-sensitive gene promoters and modulates transcription (65,66,68,74). The growing family of 20 or more NF-κB and I-kB genetic variants adds a further dimension of complexity to NF-κB and I-kB regulation and brain genetic circuitry under NF-κB control (55,66,68). Because the NF-κB complex exists as a latent cytoplasmic TF with regulated activity, it provides a rapid signaling link between extracellular events, the cytoplasm, and the nucleus, typically on the order of minutes (55,68,75). NF-κB co-localized to postsynaptic densities further suggests a role for this TF in retrograde signal transmission from the synapse to the nucleus (71,75,76). Interestingly, rapid NFκB-promoter DNA signaling is triggered by a remarkably diverse array of physiologic and pharmacologic ligands, particularly via ROS directly or by events induced by ROS, by ultraviolet irradiation, by viral, bacterial, and membrane or cellular catabolic products resulting from acute or chronic brain injury (68,72,74); by A β peptides (55,77,78), by neurotransmitters such as glutamate (79), by chemotactic peptides (80), by growth factors such as nerve-growth factor (81), and by diverse classes of neuroactive pharmacologic agents including salicylates and opioids (81–83). NF-kB knockout mice further illustrate the importance of NF-κB as a ROSresponsive TF in the maintenance of postmitotic neuronal function during aging; neuronal ROS production appears to be specifically involved in signal-transduction pathways that culminate in NF-κB activation and gene signaling (75,84–86). Mobilization of NF-κB occurs in

acute brain-injury models, such as in aluminum-induced neurotoxicity (33,86,87); in hypoxia and ischemia-reperfusion damage; during kainate-induced seizures (68,85,86); in cerebral infarctions (88); in neurodegenerative disorders including amyotrophic lateral sclerosis, Down's syndrome, Parkinson's disease, and AD (15,55,59,71,89); and in the activation of human viral genes associated with neurodegeneration, including human immunodeficiency virus (68–71). Acute brain injury also appears to increase the production of IL-1β and related cytokines, which, through the IL-1 type 1 receptor (72,90), then mediate rapid transcription from pro-inflammatory genes under NF-κB control (3,75). Via ROS production, prominent NF-κB targets include the promoters of genes encoding pathologic cytokines, such as TNF α , IL2, and IL6, vascular and intracellular adhesion molecules (VCAM-1 and ICAM-1), superoxide dismutase, βAPP, PS1, PS2, interferon β , interleukin-1 β precursor, cytosolic phospholipase A_2 (cPLA₂), and the inducible forms of nitric oxide synthase (NOS) and COX-2 (55,84-91).

Hypoxia-Inducible Factor

Analogous in structure, function, and mechanism of activation to NF-κB is hypoxiainducible factor-1 (HIF-1). HIF-1 is a heterodimeric DNA-binding protein consisting of a ROS-, cytokine-, growth factor-, and metal ion-induced HIF-1α and a constitutive HIF-1β/aryl hydrocarbon receptor nuclear transporter (ARNT) element, which in hypoxia promotes the expression of adaptive genes involved in angiogenesis, erythropoiesis, glycolysis, and related energy functions (92–99). During normoxia, the HIF- 1α subunit is directed to rapid ubiquitination and proteasomal degradation; however, during hypoxia HIF- 1α persists, heterodimerizes with HIF- 1β , and translocates to the nucleus, where it drives transcription of genes whose cis-regulatory domains contain the relatively rare hypoxia response element (HRE) 5'-RCGTG-3' (92–94). Non-homeostatic or runaway HIF-1α and HIF-

1 heterodimer production rapidly triggers aberrant gene expression in angiogenesis, erythropoiesis, glycolysis, and neovascularization, often with progression to a lethal phenotype (92,93,97–100). It is interesting that the inflammatory cytokine IL-1β partially mimics cellular hypoxia, leading to increased HIF-1α-DNA binding and activation of COX-2 and PS2 in human gingival and synovial fibroblasts and in rat and monkey retinal neurons (53,93–95; unpublished results from the authors' laboratory). In addition to hypoxia and IL-1 β , HIF-1 α is also activated by the divalent transition metals Co²⁺, Fe²⁺, Ni²⁺, Mn²⁺, Zn²⁺, and the trivalent neurotoxin Al³⁺ (15,33,95), and is blocked by carbon monoxide and nitric oxide, both of which are ligands for flavoheme protein (93–97). Notably, the primary oxygen sensor for the HIF-1 signal-transduction pathway appears to be a plasma membrane-bound flavoheme protein complex that involves a membrane-bound Fe²⁺-Fe³⁺/FADH-FAD redox shuttle sensitive to changes in both extracellular oxygen and intracellular ROS (93,97–100). Mitochondrial electron transport chain activity also appears to be required for activation of HIF-1 (92). HIF-1 activation is inhibited by normoxia or factor inhibiting HIF-1 (FIH-1), a transcriptional corepressor that binds to HIF-1 α and inhibits its transactivation function by recruiting histone deacetylases (98). Features of the HIF-1 signaltransduction cascade and mobilization of HIF-1-sensitive gene families are shown in Figures 1 and 2.

Inflammatory Signaling in Alzheimer's Disease

Activation of ROS, oxidative stress, and pro-inflammatory signaling initiate and accelerate neural cell degeneration in AD association neocortex and hippocampal CA1 (12,13,56,57,100–103). Inducible up-regulation of eicosanoids by COX-2 is observed in AD hippocampus and neocortex. Moreover, in transgenic mice that overexpress functional human COX-2 in hippocampal neurons, cog-

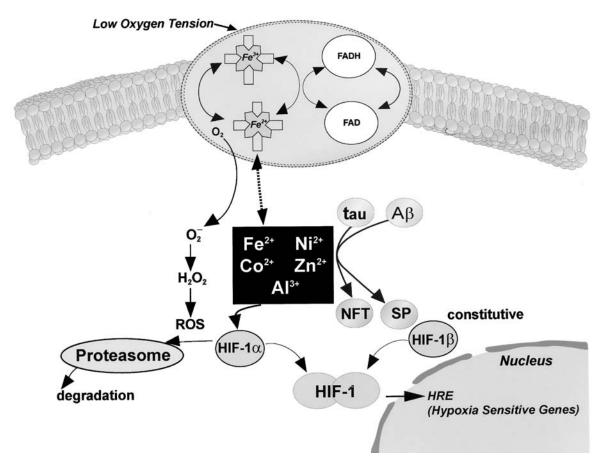


Fig. 1. Hypoxia-inducible factor-1 (HIF-1) in oxygen sensing and signaling (adapted from refs. 92–97). The primary oxygen sensors are postulated to be heme moieties localized within the plasma membrane (93,94,96,97). During normoxia, the HIF-1 α subunit undergoes rapid ubiquitination and proteasomal degradation; however, during hypoxia or pathophysiologic conditions involving prolonged periods of ischemia and/or hypoxia, HIF-1 α persists, heterodimerizes with a constitutively generated HIF-1 β , and translocates to the nucleus, where it drives transcription of hypoxia-sensitive genes whose *cis*-regulatory domains contain the relatively rare hypoxia response element (HRE) 5'-RCGTG-3' (92–94). Interestingly, Co²⁺, Fe²⁺, Ni²⁺, Mn²⁺, Zn²⁺, and the trivalent neurotoxin Al³⁺ (alone or in combination) do not require hypoxia to activate HIF-1 α . These elements also drive tau and A β aggregation into neurofibrillary tangles (NFT) and senile plaques (SP), respectively, in AD brain (93,96,97). Neurofibrillary tangles and senile plaque deposits are instrumental in proliferating the brain's inflammatory response, largely via the increased production of ROS (*see* Figure 3).

nitive deficits evolve in an age-dependent manner (102,103). PS1/PS2 holoproteins, localized to lipid raft domains in the nuclear envelope and endoplasmic reticulum, function in a nicastrin-mediated generation from β APP of neurotoxic, amyloidogenic A β pep-

tides (15,104). Importantly, transgenic animals bearing PS1/PS2 mutations or PS1/ β APP doubly transgenic animals exhibit accelerated A β generation and brain microglial activation that increase synchronously with A β deposition (105–109). The outcome of this pathologi-

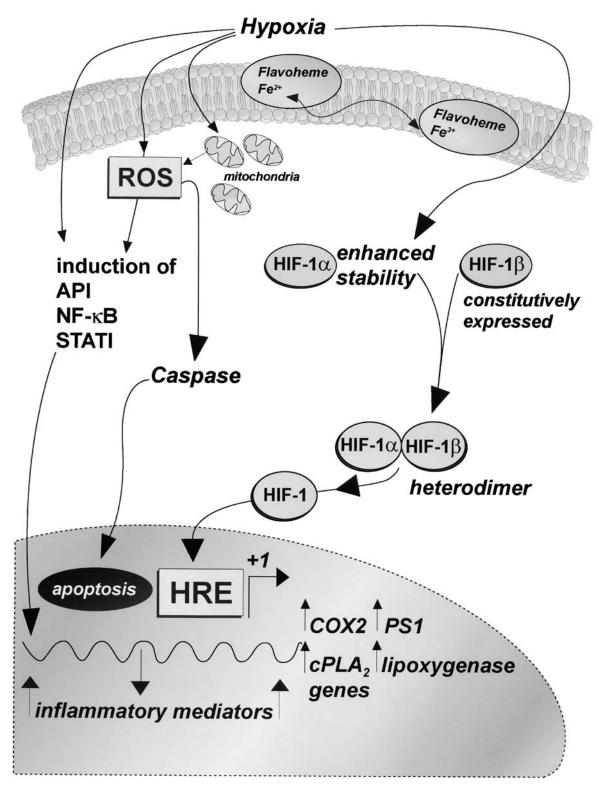


Fig. 2. Hypoxia-sensitive gene transcription. Non-homeostatic or runaway HIF-1 α and HIF-1 heterodimer production and binding to the relatively rare hypoxia response elements (HRE) in hypoxia-sensitive gene promoters rapidly trigger aberrant gene expression in angiogenesis, erythropoiesis, glycolysis, neovascularization, and neurodegeneration, often with steady progression to a lethal phenotype (92,93,97–100). ROS also induce a proinflammatory TF triad that includes AP1-, NF- κ B-, and STAT-1-DNA signaling and caspases which promote nuclear apoptosis.

cal cascade are hundred-fold increases in microglial-derived IL-1 β , related cytokines, and proinflammatory lipids, which directly correlate with increased β APP processing, generation, and aggregation of A β peptides into insoluble brain lesions, and further proliferation of COX-2 and inflammatory AD neuropathology (15,110,111).

During the microglial-mediated inflammatory response to Aβ deposition in AD, overexpression of IL-1 β not only contributes to further amyloidogenic events, but also induces a triad of the ROS-sensitive pro-inflammatory TFs: AP- 1α , NF- κ B, and STAT1 α (11,15,59,112). In the human association neocortex IL-1 β and AP1 α -, NF- κ B-, and STAT1 α -DNA binding are more abundant in AD than in age-matched control brain (55,59,111). Moreover, IL-1β-triggered AP- 1α -, NF- κ B-, and/or STAT1 α -sensitive genes, such as COX-2 and PS1, appear to have strong potential to further escalate inflammatory neuropathology. While IL-1β up-regulates the expression of COX-2 and post-translational processing of BAPP into AB42 neuropeptides, Aβ-triggered proinflammatory and neurodegenerative signaling is also induced by hypoxia, NF- κ B, and HIF-1 α . Ultimately these events trigger a hypoxia-sensitive pro-inflammatory gene cascade (8,21,112,113). These data collectively suggest that under conditions of NF- κ B and HIF-1 α activation, IL-1 β and A β 42 are further able to promote an expansion of proinflammatory pathology and an escalation of neurodegeneration (8,15,53).

Hypoxia-Triggered Gene Networks in the Brain

Phospholipase A₂-mediated AA release (as well as that of docosahexaenoic acid) and eicosanoid synthesis by COX-2 are a rapid and early response to cerebral hypoxia/ischemia. Hypoxic brain exhibits cell-membrane damage and dysfunction, membrane lipid peroxidation, ROS generation, and activation of COX-2, cytosolic and secretory phospholipase A₂ (cPLA₂, sPLA₂) activities, and lipoxygenase, which further fuel the AA

cascade (15,61). In turn, AA cascades generate inflammatory mediators and trigger cellular degeneration (Figure 3). IL-1β, transforming growth factor-beta (TGFβ) and tumor necrosis factor-alpha (TNFα), secreted by microglia, astrocytes, and neurons, are early-response cytokines that influence the progression of injury by stimulating the synthesis of other cytokines and neuronal injury mediators, such as COX-2. Therefore, via the generation of inflammatory mediators that accompany hypoxia, activation of ROS and oxygen-sensitive AP-1-, HIF-1 α -, NF- κ B-, and STAT-1 α -DNA binding trigger COX-2, PS1, and related gene expression, leading to progressive neural dysfunction. Hypoxia-modulated pathologic gene induction is therefore remarkable in that positive feedback by ROS and oxygensensitive TFs further recruits COX-2, PS1, cPLA₂, and lipoxygenase gene signaling that propagates and accelerates neurodegeneration (15,27).

COX-2 and PS1 are significant players in the sequence of events leading to the expansion and acceleration of inflammatory neuropathology (8,15,27). Because COX-2 can up-regulate eicosanoid production, and PS1 mediates Aβ generation, these in turn trigger microglial activation, the further release of IL-1 β , and COX-2 and PS1 gene expression. Importantly, COX-2 and PS1 up-regulation is observed in IL-1β- and Aβ42-triggered human neural cells in culture, and this upregulation is further increased after brief periods of exposure to hypoxia (8,20–22). The participation of ROS in inflammatory gene recruitment is a key factor, since both COX-2 and PS1 activation can be blocked by the electron spin-trap agent and free radical scavenger α -phenyl-*N*-tert-butyl nitrone (8,21,99). Interestingly, DNA microarray analysis confirms that COX-2 and PS1, as well as the NFkBp50/p105, IL-1β precursor, and cPLA₂ genes, are induced both in AD and in IL-1βand Aβ42-induced human neural cells, especially after exposure to hypoxia (21,52–54). Indeed, chromosomally unlinked, inducible TATA-box-containing genes under coregu-

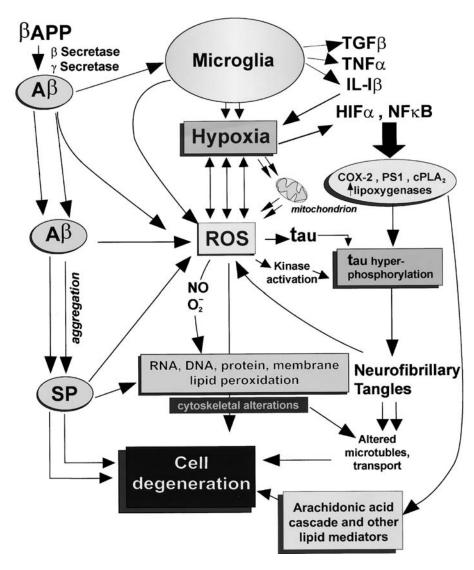


Fig. 3. Hypoxia triggers neuronal-cell degeneration. Pathways involving Aβ production and senile plaque (SP) formation are central to microglial-cytokine release, hypoxia, ROS generation, RNA, DNA, protein, and brain-lipid peroxidation. Microglial-derived cytokines proliferate hypoxia and ROS generation, which in turn, drive inflammatory AA cascades, tau hyper-phosphorylation, NFT generation and altered neural cytoarchitecture, resulting in cellular dysfunction and neurodegeneration.

lated transcriptional control by the ROS-sensitive TFs AP-1, NF- κ Bp50/p65, STAT1 α , and HIF-1 α may define, in part, a brain-specific inflammatory gene family that contribute cumulatively to pro-inflammatory neuropathology in AD and related neurodegenerative disorders (8,15).

Hypoxia, Bcl2, p53 and Apoptosis

As in the case of the COX-2, PS1, NF-kBp50/p105, IL-1 β precursor and cPLA₂ genes, signal-transduction pathways that direct cellular fate toward apoptosis and/or necrosis and neural degeneration appear to be

modulated in part by HIF-1α- and NF-κB-DNA binding in target gene promoters (8,15,21,46,52–56,59–62). Hypoxia also induces NF- κ B binding to the Bcl-x promoter, activating transcription and alternate RNA splicing to the Bcl-XS transcript variant that promotes hippocampal neuronal apoptosis (114,115). Similarly, induction of HIF- 1α -DNA binding by hypoxia induces PS2, a regulator of intercellular signaling during CNS development and cell-fate determination that is involved in βAPP processing into neurotoxic Aβ peptides (95,96). Intracellular A β peptides are cytotoxic to human neurons in part through the p53-Bax cell death-signaling pathways (28,115,116), hence PS2 gene activation during hypoxia may direct cell metabolism towards intercellular PS2-mediated signaling dysfunction. The p53 tumor suppressor that normally limits cellular proliferation by arresting cell-cycle progression also induces apoptosis as a response to hypoxia (28,99,117,118). p53 is thought to mediate apoptosis through Bax transactivation, translocation from the cytosol to membranes, triggering of cytochrome c release from mitochondria, and activation of proteolytic caspases (115,117). Interestingly, both wild-type and mutated PS2 appear to trigger p53-mediated apoptotic cascades, both in HEK293 human cells and in murine neurons (119), suggesting a rapid and immediate link between hypoxia-activated PS2 gene expression and neural apoptosis (92–99).

Summary

The etiopathogenesis of AD is multi-factorial and highly complex. This review suggests key roles for hypoxia in initiating, regulating, and proliferating inflammatory responses and determining cell-fate pathways in the brain. Abundant data from neonatal, developing, aging, and degenerating brain tissue suggest an early involvement of oxidative stress, via ROS, in the pathogenesis of AD. This may provide potential targets for therapeutic intervention,

especially in individuals of families at high genetic risk for developing AD. These data also suggest that hypoxia over the course of aging may drive inflammatory gene programs via complementary, interdependent TF signaling pathways, and these may contribute cumulatively to inflammatory signaling and brain-cell degeneration. Because these inflammatory activation mechanisms can be quenched by free radical scavengers, diets enriched in antioxidants may help to repress the potential inception and development of AD and related neurodegenerative conditions.

Acknowledgment

The authors thank Aileen Pogue, Darlene Guillot, and Carole Lachney for expert editorial assistance. This work was supported in part by United States Public Health Service grants AG18031 from the National Institute on Aging (to WJL), NS23002 from the National Institute of Neurologic Disorders and Stroke (to NGB), and by the Eye, Ear, Nose and Throat Foundation of Louisiana.

References

- 1. Oka A. and Takashima S. (1997) Induction of cyclooxygenase 2 in brains of patients with Down's syndrome and dementia of Alzheimer type: specific localization in affected neurones and axons. *Neuroreport.* **24**, 1161–1164.
- 2. Lukiw W. J. and Bazan N. G. (1997) Cyclooxygenase 2 RNA message abundance, stability, and hypervariability in sporadic Alzheimer neocortex *J. Neurosci. Res.* **50**, 937–945.
- 3. Perkins D. J. and Kniss D. A. (1997) Tumor necrosis factor-alpha promotes sustained cyclooxygenase-2 expression: Attenuation by dexamethasone and NSAIDS. *Prostaglandins.* **54**, 727–743.
- Tateishi N., Kagamiishi Y., Shintaku K., Shimoda T., Shinagawa R., Satoh S., and Kondo K. (1998). Activation of astrocytes and ischemic damage following the transient focal ischemia. *Nippon Yakurigaku Zasshi* 112, S1, 103P–107P.

- Tanimukai H., Imaizumi K., Kudo T., Katayama T., Tsuda M., Takagi T., Tohyama M., and Takeda M. (1998) Alzheimer-associated presenilin-1 gene is induced in gerbil hippocampus after transient ischemia. *Brain Res. Mol. Brain Res.* 54, 212–218.
- Miake H., Tsuchiya K., Nakamura A., Ikeda K., Levesque L., Fraser P. E., St-George Hyslop P. H., Mizusawa H., and Uchihara T. (1999) Glial expression of presenilin epitopes in human brain with cerebral infarction and in astrocytoma. *Acta Neuropathol.* (Berl.) 98, 337–340.
- 7. Pennypacker K. R., Hernandez H., Benkovic S., Morgan D. G., Willing A. E., and Sanberg P. R. (1999) Induction of presentilins in the rat brain after middle cerebral arterial occlusion. *Brain Res. Bull.* **48**, 539–543.
- 8. Bazan N. G. and Lukiw W. J. (2002) COX-2 and PS1 Gene Expression Induced by IL-1β and Aβ42 is Potentiated by Hypoxia in Primary Human Neural Cells. *J. Biol. Chem.* **277**, 30,359–30,367.
- 9. Scheff S. W. and Price D. A. (1998) Synaptic density in the inner molecular layer of the hippocampal dentate gyrus in Alzheimer disease. *J. Neuropathol. Exp. Neurol.* **57**, 1146–1153.
- 10. Yasojima K., Schwab C., McGeer E. G., and McGeer P. L. (1999) Distribution of cyclooxygenase-1 and cyclooxygenase-2 mRNAs and proteins in human brain and peripheral organs. *Brain Res.* **830**, 226–236.
- 11. Akama K. T. and Van Eldik L. J. (2000) Betaamyloid stimulation of inducible nitric-oxide synthase in astrocytes is interleukin-1beta- and tumor necrosis factor-alpha (TNFalpha)dependent, and involves a TNFalpha receptorassociated factor- and NFkappaB-inducing kinase-dependent signaling mechanism. *J. Biol. Chem.* **275**, 7918–7924.
- 12. Combs C. K., Johnson D. E., Karlo J. C., Cannady S. B., and Landreth G. E. (2000) Inflammatory mechanisms in Alzheimer's disease: inhibition of beta-amyloid-stimulated proinflammatory responses and neurotoxicity by PPARgamma agonists. *J. Neurosci.* **20**, 558–567.
- 13. Rogers J. and Shen Y. (2000) A perspective on inflammation in Alzheimer's disease. *Ann. NY. Acad. Sci.* **924**, 132–135.
- 14. Czech C., Tremp G., and Pradier L. (2000) Presenilins and Alzheimer's disease: biological functions and pathogenic mechanisms. *Prog. Neurobiol.* **60**, 363–384.

- 15. Lukiw W. J. and Bazan N. G. (2000) Neuroinflammatory signaling upregulation in Alzheimer's disease. *Neurochem. Res.* **25**, 1173–1184.
- 16. Selkoe D. J. (2000) Notch and presenilins in vertebrates and invertebrates: implications for neuronal development and degeneration. *Curr. Opin. Neurobiol.* **10**, 50–57.
- 17. Selkoe D. J. (2001) Presenilin, Notch, and the genesis and treatment of Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* **98**, 11,039–11,041.
- 18. Cummings J. L. and Cole G. (2002) Alzheimer disease. *JAMA* **287**, 2335–2338.
- 19. Hull M., Lieb K. and Fiebich B. L. (2002) Pathways of inflammatory activation in Alzheimer's disease: potential targets for disease modifying drugs. *Curr. Med. Chem.* **9**, 83–88.
- 20. Pratico D. (2002) Alzheimer's disease and oxygen radicals: new insights. *Biochem. Pharmacol.* **63**, 563–567.
- 21. Butterfield D. A, and Lauderback C. M. (2002) Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and consequences involving amyloid betapeptide-associated free radical oxidative stress. *Free Radic. Biol. Med.* 32, 1050–1060.
- 22. Love S. (1999). Oxidative stress in brain ischemia. *Brain Pathol.* **9,** 119–131.
- 23. Bonazzi A., Mastyugin V., Mieyal P. A., Dunn M. W., and Laniado-Schwartzman M. (2000) Regulation of cyclooxygenase-2 by hypoxia and peroxisome proliferators in the corneal epithelium. *J. Biol.Chem.* **275**, 2837–2844.
- 24. Carter J., and Lippa C. F. (2001) Beta-amyloid, neuronal death and Alzheimer's disease. *Curr. Mol. Med.* **6**, 733–737.
- 25. Holmes C. (2002) Genotype and phenotype in Alzheimer's disease. *Br. J. Psychiatry* **180,**131–134.
- DeKosky S. T. (2001) Epidemiology and pathophysiology of Alzheimer's disease. *Clin. Cor*nerstone 3, 15–26.
- 27. Bazan N. G. (2001) COX-2 as a multifunctional neuronal modulator. *Nat. Med.* **7**, 414–415.
- 28. Mattson M. P., Duan W., Pedersen W. A., and Culmsee C. (2001) Neurodegenerative disorders and ischemic brain diseases. *Apoptosis* **6**, 69–81.
- 29. Marshall L. F. (2000) Head injury: recent past, present, and future. *Neurosurgery* **47**, 546–561.
- 30. Desmond D. W. (1996) Vascular dementia: a construct in evolution. *Cerebrovasc. Brain Metab. Rev.* 8, 296–325.

- 31. Gitter B. D., Boggs L. N., May P. C., Czilli D. L., and Carlson C. D. (2000) Regulation of cytokine secretion and amyloid precursor protein processing by proinflammatory amyloid beta (Aβ). *Ann. NY Acad . Sci.* **917**, 154–164.
- 32. Rhodin J., Thomas T., Bryant M., and Sutton E. T. (2000) Animal model of Alzheimer-like vascular pathology and inflammatory reaction. *Ann. NY Acad. Sci.* **903**, 345–352.
- 33. Campbell A., Yang E. Y., Tsai-Turton M., and Bondy S. C. (2002) Pro-inflammatory effects of aluminum in human glioblastoma cells. *Brain Res.* **933:** 60–65.
- 34. John G. R., Simpson J. E., Woodroofe M. N., Lee S. C, and Brosnan C. F. (2001) Extracellular nucleotides differentially regulate interleukin-1beta signaling in primary human astrocytes: implications for inflammatory gene expression. *J. Neurosci.* 21, 4134–4142.
- 35. Apelt J. and Schliebs R. (2001) Beta-amyloid-induced glial expression of both pro- and anti-inflammatory cytokines in cerebral cortex of aged transgenic Tg2576 mice with Alzheimer plaque pathology. *Brain Res.* **894**, 21–30.
- 36. Fortini M. E. (2001) Notch and presenilin: a proteolytic mechanism emerges. *Curr. Opin. Cell. Biol*. **13**, 627–634.
- 37. Blass J. P. (2001) Brain metabolism and brain disease: is metabolic deficiency the proximate cause of Alzheimer dementia? *J. Neurosci. Res.* **66:** 851–856.
- 38. Kaufmann W. A., Barnas U., Humpel C., Nowakowski K., DeCol C., Gurka P., Ransmayr G., Hinterhuber H., Winkler H., and Marksteiner J. (1998) Synaptic loss reflected by secretoneurin-like immunoreactivity in the human hippocampus in Alzheimer's disease. *Eur. J. Neurosci.* **10**, 1084–1094.
- 39. Yao P. J., Morsch R., Callahan L. M., and Coleman P. D. (1999) Changes in synaptic expression of clathrin assembly protein AP180 in Alzheimer's disease analysed by immunohistochemistry. *Neuroscience* **94**, 389–394.
- 40. Roses A. D. (1997) Apolipoprotein E, a gene with complex biological interactions in the aging brain. *Neurobiol. Dis.* **4,** 170–185.
- 41. Neve R. L. and Robakis N. K. (1998) Alzheimer's disease: a re-examination of the amyloid hypothesis. *Trends Neurosci.* **21**, 15–19.
- 42. St. George-Hyslop P. H. (2000) Molecular genetics of Alzheimer's disease. *Biol. Psychiatry* 47, 183–199.

- 43. McDowell I. (2002). Alzheimer's disease: insights from epidemiology. *Aging (Milano)* **13**, 143–162.
- 44. Daniels W. M., Hendricks J., Salie R., and Taljaard J. J. (2001) The role of the MAP-kinase superfamily in beta-amyloid toxicity. *Metab. Brain Disease* **16**, 175–185.
- 45. Holcomb L., Gordon M. N., McGowan E., Yu X., Benkovic S., Jantzen P., Wright K., Saad I., Mueller R., Morgan D., Sanders S., Zehr C., O'Campo K., Hardy J., Prada C. M., Eckman C., Younkin S., Hsiao K., and Duff K. (1998) Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes. *Nat. Med.* 4, 97–100.
- 46. Lukiw W. J., Rogaev E. I., and Bazan N. G. (1996) Synaptic and cytoskeletal RNA message levels in sporadic Alzheimer neocortext. *Alzheimers Res.* **2**, 221–227.
- 47. Chow N., Cox C., Callahan L. M., Weimer J. M., Guo L., and Coleman P. D. (1998) Expression profiles of multiple genes in single neurons of Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* **95**, 9620–9625.
- Tol J., Roks G., Slooter A. J., and van Duijn C. M. (1999) Genetic and environmental factors in Alzheimer's disease *Rev. Neurol.* 155, 10–16.
- 49. Lukiw W. J., Rogaev E. I., Wong L., Vaula G., McLachlan D. R., and St. George- Hyslop P. (1994) Protein-DNA interactions in the promoter region of the amyloid precursor protein (APP) gene in human neocortex . *Mol .Brain. Res.* 22, 121–131.
- 50. Van Leeuwen F., Wde Kleijn D. P., van den Hurk H. H., Neubauer A., Sonnemans M. A., Sluijs J. A., Koycu S., Ramdjielal R. D. J., Salehi A., Martens G. J. M., Grosveld F. G., Peter J., Burbach H., and Hol E. M. (1998) Frameshift mutants of beta amyloid precursor protein and ubiquitin-B in Alzheimer's and Down patients. *Science* 279, 242–247.
- 51. Rogaev E. I., Sherrington R., Wu C., Levesque G., Liang Y., Rogaeva E. A., Ikeda M., Holman K., Lin C, Lukiw W. J., de Jong P. J., Fraser P. E., Rommens J. M., and St. George-Hyslop P. (1997) Analysis of the 5' sequence, genomic structure, and alternative splicing of the presenilin-1 gene (PSEN1) associated with early onset Alzheimer disease. *Genomics* 40, 415–424.
- 52. Lukiw W. J., Carver C. A., LeBlanc H. J., and Bazan N. G. (2000) Analysis of 1184 gene transcript levels in Alzheimer CA1 hippocampus;

- synaptic signaling and transcription factor deficits and up-regulation of pro-inflammatory pathways. *Alz. Rep.* **3**, 161–167.
- 53. Lukiw W. J., Ottlecz A., Lambrou G., Grueninger M., Finley J., and Bazan N. G. (2002) Activation of HIF-1α and NF-kB-DNA Binding and COX-2 and VEGF Gene Expression in Monkey Choroid-Retinal (RF/6A) Cells by Hypoxia . *Invest. Ophthalmol. Vis. Sci.* **42:** S243.
- 54. Loring J. F., Wen X., Lee J. M., Seilhamer J., and Somogyi R. A. (2002) Gene expression profile of Alzheimer's disease. *DNA Cell Biol.* **20**: 683–695.
- 55. Lukiw W. J. and Bazan N. G. (1998) Strong nuclear factor-kappaB-DNA binding parallels cyclooxygenase-2 gene transcription in aging and in sporadic Alzheimer's disease superior temporal lobe neocortex. *J. Neurosci. Res.* 1, 583–592.
- 56. McGeer E. G. and McGeer P. L. (1999) Brain inflammation in Alzheimer disease and the therapeutic implications. *Curr. Pharm. Res.* **10**, 821–836.
- 57. McGeer P. L. McGeer, E. G., and Yasojima K. J. (2000) Alzheimer disease and neuroinflammation. *Neural Transm. Suppl.* **59**, 53–57.
- 58. Ho L., Pieroni C., Winger D., Purohit D. P., Aisen P. S., and Pasinetti G. M. (1999) Regional distribution of cyclooxygenase-2 in the hippocampal formation in Alzheimer's disease. *J. Neurosci. Res.* 57, 295–303.
- 59. Kitamura Y., Shimohama, S., Ota T., Matsuoka Y., Nomura Y., and Taniguchi T. (1997) Alteration of transcription factors NF-kappaB and STAT1 in Alzheimer's disease brains. *Neurosci. Lett.* **237**, 17–20.
- 60. Kitamura Y., Shimohama S., Koike H., Kakimura J., Matsuoka Y., Nomura, Y., Gebicke-Haerter P. J., and Taniguchi T. (1999) Increased expression of cyclooxygenases and peroxisome proliferator-activated receptor-gamma in Alzheimer's disease brains. *Biochem. Biophys. Res. Commun.* **254**, 582–586.
- 61. Mishra O. P. and Delivoria-Papadopoulos M. (1999) Cellular mechanisms of hypoxic injury in the developing brain. *Brain Res. Bull.* **48**, 233–8.
- 62. Aliev G., Smith M. A., Seyidov D., Neal M. L., Lamb B. T., Nunomura A., Gasimov E. K., Vinters H. V., Perry G., LaManna J. C., and Friedland R. P. (2002) The role of oxidative stress in the pathophysiology of cerebrovascular lesions in Alzheimer's disease. *Brain Pathol.* 12, 21–35.

- 63. Nakamura Y., Nakashima T., Fukuda S., Nakashima H., and Hashimoto T. (1986) Hypoxic-ischemic brain lesions found in asphyxiating neonates. *Acta Pathol.* (*Japan*) **36**, 551–563.
- 64. Tsang M. C., Lo A. C., Cheung P. T., Chung S. S., and Chung S. K. Perinatal hypoxia-/ischemia-induced endothelin-1 mRNA in astrocyte-like and endothelial cells. *Neurore-port* 2001 **12**, 2265–2270.
- 65. Sen R. and Baltimore D. (1986) Multiple nuclear factors interact with the immunoglobulin enhancer sequences. *Cell.* **46,** 705–716.
- 66. Baeuerle P. A. and Baltimore D. (1996) NF-κB: Ten years after. *Cell.* **87**, 13–20.
- 67. Lukiw W. J., Pelaez R. P., Martinez J., and Bazan N. G. (1998) Budesonide epimer R or dexamethasone selectively inhibit platelet-activating factor-induced or interleukin 1beta-induced DNA binding activity of cis-acting transcription factors and cyclooxygenase-2 gene expression in human epidermal keratinocytes. *Proc. Natl. Acad. Sci. USA* **95**, 3914–3919.
- 68. Li X. and Stark G. R. (2002). NFkappaB-dependent signaling pathways. *Exp. Hematol.* **30**, 285–96.
- 69. Kaltschmidt C., Kaltschmidt B., Neumann H., Wekerle H., and Baeuerle P. A. (1994) Constitutive NF-kappa B activity in neurons. *Mol. Cell Biol.* **14**, 3981–3992.
- 70. Baeuerle P. A. and Henkel T. (1994) Function and activation of NF-kappa B in the immune system. *Annu. Rev. Immunol.* **12**, 141–179.
- 71. Kaltschmidt B., Uherek M., Volk B., Bacuerle P. A., and Katlschmidt C. (1997) Transcription factor NF-kappa B is activated in primary neurons by amyloid beta peptides and in neurons surrounding early plaques from patients with Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 94, 2642–2647.
- 72. Stylianou E., O'Neill L. A., Rawlinson L., Edbrooke M. R., Woo P., and Saklatvala J. (1992) Interleukin 1 induces NF-kappa B through its type I but not its type II receptor in lymphocytes. *J. Biol. Chem.* **267**, 15836–15841.
- 73. Traenckner E. B., Pahl H. L., Henkel T., Schmidt K. N., Wilk S., and Baeuerle P. A. (1995) Phosphorylation of human I kappa Balpha on serines 32 and 36 controls I kappa Balpha proteolysis and NF-kappa Bactivation in response to diverse stimuli. *EMBO J.* 14, 2876–2883.

- 74. Verma I. M., Stevenson J. K., Schwarz E. M., Van Antwerp D., and Miyamoto S. (1995) Rel/NF-kappa B/I kappa B family: Intimate tales of association and dissociation. *Genes Dev.* **15**, 2723–2735.
- 75. Mattson M. P. and Camandola S. (2001). NF-kappaB in neuronal plasticity and neurodegenerative disorders. *J. Clin. Invest.* **107**, 247–254.
- 76. Suzuki T., Mitake S., Okumura-Noji K., Yang J. P., Fujii T., and Okamoto T. (1997) Presence of NF-kappa B-like and I kappa B-like immunoreactivities in postsynaptic densities. *Neuroreport.* **8**, 2931–2935.
- Akama K. T., Albanese C., Pestell R. G., and Van Eldik L. J. (1998) Amyloid beta-peptide stimulates nitric oxide production in astrocytes through an NFkappaB-dependent mechanism. *Proc. Natl. Acad. Sci. USA* 95, 5795–5800.
- 78. Kuner P., Schubenel R., and Hertel C. (1998) Beta-amyloid binds to p57NTR and activates NFkappaB in human neuroblastoma cells. *J. Neurosci. Res.* **54**, 798–804.
- 79. Guerrini L., Molteni A., Wirth T., Kistler B., and Blasi F. (1997) Glutamate-dependent activation of NF-kappa B during mouse cerebellum development. *J. Neurosci.* 17, 6057–6063.
- 80. Browning D. D., Pan Z. K., Prossnitz E. R., and Ye R. D. (1997) Cell type- and developmental stage-specific activation of NF-kappa B by fMet-Leu-Phe in myeloid cells. *J. Biol. Chem.* **272**, 7995–8001.
- 81. Carter B. D., Kaltschmidt C., Kaltschmidt B., Offenhauser N., Bohm-Matthaei R., Baeuerle P. A., and Barde Y. A. (1996) Selective activation of NF-kappa B by nerve growth factor through the neurotrophin receptor p75. *Science*. 272, 542–545.
- 82. Cai W., Hu L., and Foulkes J. G. (1996) Transcription-modulating drugs: Mechanism and selectivity. *Curr. Opin. Biotechnol.* **7**, 608–615.
- 83. Hou Y. M., Vlaskovska M., Cebers G., Kasakov L., Liljequist S., and Terenius L. (1996) A mureceptor opioid agonist induces AP-1 and NF-kappa B transcription factor activity in primary cultures of rat cortical neurons. *Neurosci. Lett.* **212**, 159–162.
- 84. Schmidt-Ullrich R., Memet S., Lillenbaum A., Feuillard J., Raphael M., and Israel A. (1996) NF-kappa B activity in transgenic mice: Developmental regulation and tissue specificity. *Development.* **122**, 2117–2128.
- 85. Rong Y. and Baudry M. (1996) Seizure activity results in a rapid induction of nuclear factor-

- kappa B in adult but not juvenile rat limbic structures. *J. Neurochem.* **67**, 662–668.
- 86. O'Neill L. A. and Kaltschmidt C. (1997) NF-kappa B: A crucial transcription factor for glial and neuronal cell function. *Trends Neurosci.* **20**, 252–258.
- 87. Mundy W. R., Freudenrich T. M., and Kodavanti P. R. (1997) Aluminum potentiates glutamate-induced calcium accumulation and iron-induced oxygen free radical formation in primary neuronal cultures. *Mol. Chem. Neuropathol.* **32**, 41–57.
- 88. Terai K., Matsuo A., McGeer E. G., and McGeer P. L. (1996) Enhancement of immunoreactivity for NF-kappa B in human cerebral infarctions. *Brain Res.* **739**, 343–349.
- 89. Hunot S., Brugg B., Ricard D., Michel P. P., Muriel M. P., Ruberg M., Faucheux B. A., Agid Y., and Hisch E. C. (1997) Nuclear translocation of NF-kappa B is increased in dopaminergic neurons of patients with Parkinson disease. *Proc. Natl. Acad. Sci. USA.* **94**, 7531–7536.
- 90. Lukiw W. J., Martinez J., Pelaez R. P., and Bazan N. G. (1999) The interleukin-1 type 2 receptor gene displays immediate early gene responsiveness in glucocorticoid-stimulated human epidermal keratinocytes. *J. Biol. Chem.* **274**, 8630–8638.
- 91. Yamamoto Y. and Gaynor R. B. (2001) Role of the NF-kappaB pathway in the pathogenesis of human disease states. *Curr. Mol. Med.* 1, 287–296.
- 92. Agani F. H., Pichiule P., Chavez J. C., and LaManna J. C. (2000) The role of mitochondria in the regulation of hypoxia-inducible factor 1 expression during hypoxia. *J. Biol. Chem.* **275**, 35,863–35,867.
- 93. Semenza, G. L. (2001) Hypoxia-inducible factor 1: oxygen homeostasis and disease pathophysiology. *Trends Mol. Med.* 7, 345–350.
- 94. Thornton R. D., Lane P., Borghaei R. C., Pease E. A., Caro J., and Mochan E. (2000) Interleukin 1 induces hypoxia-inducible factor 1 in human gingival and synovial fibroblasts. *J. Biochem.* **350**, Pt 1, 307–312.
- 95. Lukiw, W. J., Gordon W. C., Rogaev E. I, Thompson H., and Bazan N. G. (2001a) Presenilin-2 (PS2) expression up-regulation in a model of retinopathy of prematurity and pathoangiogenesis. *Neuroreport* **12**, 53–57.
- 96. Lukiw W. J. (2001) Aluminum and gene transcription in the mammalian central nervous system in *Aluminum and Alzheimer's Disease*,

- *The Science That Describes the Link* (Edited by C. Exley) Elsevier Press, New York, pp.147–168.
- 97. Zhu H. and Bunn H. F. (1999) Oxygen sensing and signaling: impact on the regulation of physiologically important genes. *Respir. Physiol.* **115**, 239–247.
- 98. Mahon P. C., Hirota K., Semenza G. L.. (2001) FIH-1: a novel protein that interacts with HIF-1alpha and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev* **15**, 2675–2686.
- 99. Zhu H., Jackson T., and Bunn H. F. (2002) Detecting and responding to hypoxia. *Nephrol. Dial. Transplant* **17**, Suppl 1:3–7.
- 100. Butterfield D. A., Howard B., Yatin S., Koppal T., Drake J., Hensley K., Aksenov M., Aksenova M., Subramaniam R., Varadarajan S., Harris-White M. E., Pedigo N. W. Jr., and Carney J. M. (1999) Elevated oxidative stress in models of normal brain aging and Alzheimer's disease. *Life Sci.* 65, 1883–1892.
- 101. Rothwell N. J. and Luheshi G. N. (2000) Interleukin 1 in the brain: biology, pathology and therapeutic target. *Trends Neurosci.* 23, 618–625.
- 102. Kaufmann W. E., Andreasson K. I., Isakson P. C., and Worley P. F. (1997) Cyclooxygenases and the central nervous system. *Prostaglandins*. **54**, 601–624.
- 103. Andreasson K. I., Savonenko A., Vidensky S., Goellner J. J., Zhang Y., Shaffer A., Kaufmann W. E., Worley P. F., Isakson P., and Markowska A. L. (2001) Age-dependent cognitive deficits and neuronal apoptosis in cyclooxygenase-2 transgenic mice. *J. Neurosci.* 21, 8198–8209.
- 104. Matsuoka Y., Picciano M., Malester B., LaFrancois J., Zehr C., Daeschner J. M., Olschowka J. A., Fonseca M. I., O'Banion M. K., Tenner A. J., Lemere C. A., and Duff K. (2001) Inflammatory responses to amyloidosis in a transgenic mouse model of Alzheimer's disease. *Am. J. Pathol.* **158**, 1345–1354.
- 105. Niwa M., Sidrauski C., Kaufman R. J., and Walter P. (1999) A role for presenilin-1 in nuclear accumulation of Ire1 fragments and induction of the mammalian unfolded protein response. *Cell.* **99**, 691–702.
- 106. Annaert W. G., Levesque L., Craessaerts K., Dierinck I., Snellings G., Westaway D., George-Hyslop P. S., Cordell B., Fraser P., and De Strooper B. (1999) Presenilin 1 controls gamma-secretase processing of amyloid precursor protein in pre-golgi compartments of hippocampal neurons. J. Cell. Biol. 147, 277–294.

- 107. Honda T., Nihonmatsu N., Yasutake K., Ohtake A., Sato K., Tanaka S., Murayama O., Murayama M., and Takashima A. (2000) Familial Alzheimer's disease-associated mutations block translocation of full-length presenilin 1 to the nuclear envelope. *Neurosci. Res.* 37, 101–111.
- 108. Wiltfang J., Esselmann H., Cupers P., Neumann M., Kretzschmar H., Beyermann M., Schleuder D., Jahn H., Ruther E., Kornhuber J., Annaert W., De Strooper B., and Saftig P. (2001) Elevation of beta-amyloid peptide 2-42 in sporadic and familial Alzheimer's disease and its generation in PS1 knockout cells. *J. Biol. Chem.* **276**, 42,645–42,657.
- 109. Dewachter I., Moechars D., van Dorpe J., Tesseur I., Van den Haute C., Spittaels K., and Van Leuven F. (2001) Modelling Alzheimer's disease in multiple transgenic mice. *Biochem. Soc. Symp.* **67**, 203–210.
- 110. Goldgaber D., Harris H. W., Hla T., Maciag T., Donnelly R. J., Jacobsen J. S., Vitek M. P., and Gajdusek D. C. (1989) Interleukin 1 regulates synthesis of amyloid beta-protein precursor mRNA in human endothelial cells. *Proc. Nat. Acad. Sci. USA* **86**, 7606–7610.
- 111. Griffin W. S., Nicoll J. A., Grimaldi L. M., Sheng J. G., and Mark R. E. (2000) The pervasiveness of interleukin-1 in alzheimer pathogenesis: a role for specific polymorphisms in disease risk. *Exp. Gerontol.* **35**, 481–487.
- 112. Hoozemans J. J., Veerhuis R., Janssen I., Rozemuller A., and Eikelenboom P. (2001) Interleukin-1beta induced cyclooxygenase 2 expression and prostaglandin E2 secretion by human neuroblastoma cells: implications for Alzheimer's disease. *Exp. Gerontol.* **36**, 559–570.
- 113. Fiebich B. L., Mueksch B., Boehringer M., and Hull M. (2000) Interleukin-1beta induces cyclooxygenase-2 and prostaglandin E(2) synthesis in human neuroblastoma cells: involvement of p38 mitogen-activated protein kinase and nuclear factor-kappaB. *J. Neurochem.* 275, 2020–2028.
- 114. Glasgow J. N., Qiu J., Rassin D., Grafe M., Wood T., and Perez-Polo J. R. (2001) Transcriptional regulation of the BCL-X gene by NF-kappaB is an element of hypoxic responses in the rat brain. *Neurochem. Res.* **26**, 647–659.
- 115. Zhang Y., McLaughlin R., Goodyer C., and LeBlanc A. (2002) Selective cytotoxicity of intracellular amyloid beta peptide 1-42

- through p53 and Bax in cultured primary human neurons. *J. Cell Biol.* **156,** 519–529.
- 116. Mattson M. P., Zhu H., Yu J., and Kindy M. S. (2000) Presenilin-1 mutation increases neuronal vulnerability to focal ischemia in vivo and to hypoxia and glucose deprivation in cell culture: involvement of perturbed calcium homeostasis. *J. Neurosci.* **20**, 1358–1364.
- 117. Shen Y. and White E. (2001) p53-dependent apoptosis pathways *Adv Cancer Res* **82**, 55–84.
- 118. Jones M. K., Szabo I. L., Kawanaka H., Husain S. S., Tarnawski A. S., and von Hippel Lindau

- (2002) Tumor suppressor and HIF-1alpha: new targets of NSAIDs inhibition of hypoxia-induced angiogenesis. *FASEB J.* **16**, 264–266.
- 119. Alves da Costa C., Paitel E., Mattson M. P., Amson R., Telerman A., and Ancolio K., Checler F. (2002) Wild-type and mutated presenilins 2 trigger p53-dependent apoptosis and down-regulate presenilin 1 expression in HEK293 human cells and in murine neurons. *Proc. Natl. Acad. Sci. USA* **99**, 4043–4048.